β₂ Microglobulin as A Marker of Early Renal Damage In Patients With Sickle Cell Nephropathy

Dr. Alaje Abiodun K¹, Dr. Nwogoh Benedict², Idogun Sylvester E³.

¹Department of Chemical Pathology, University of Benin Teeaching Hospital, Benin City. ²Department of Haematology, University of Benin Teaching Hospital, Benin City. ³Department of Chemical Pathology, University of Benin Teeaching Hospital, Benin City.

Abstract

Introduction: Sickle cell nephropathy is a cause of significant morbidity and mortality in patients with sickle cell disease. Traditional creatinine based assay for evaluating renal function has several limitations. Hence, the need for more sensitive markers such as β_2 microglobulin ($\beta_2 M$).

This study seeks to compare $\beta_2 M$ with creatinine as a marker of renal damage in patients with sickle cell disease.

Method: This is a cross sectional study conducted at the University of Benin Teaching Hospital, Benin City, Nigeria. Serum creatinine and $\beta_2 M$ levels were determine using kinetic Jaffe method and $\beta_2 M$ ELISA assay (Quantikine kit) respectively in 83 sickle cell disease patients and 40 age and sex match controls. Estimated glomerular filtration rate (eGFR) was calculated using Cockcroft-Gault formular for eGFRCr estimation. Data was analyzed with SPSS version 16.

Result: $\beta_2 M$ in the SCD patients and controls were $25.0\pm3.9\%$ versus $45.6\pm5.8\%$ (p = 0.001), $17.4\pm7.5mg/dl$ versus $24.0\pm8.0mg/dl$ (0.001), $0.7\pm0.3mg/dl$ versus $0.9\pm0.3mg/dl$ (p = 0.003), 3.3 ± 1.0 versus 2.3 ± 0.7 (p = 0.001) respectively. The eGFR-Cr was higher in SCD patients than in controls ($128.9\pm8.4ml/min/1.73m^2$ versus $113.6\pm11.0ml/min/1.73m^2$) but the difference was not statistically significant P = 0.316. $\beta_2 M$ has a positive correlation with urea, creatinine and a weak negative correlation with eGFRCr.

Conclusion: $\beta_2 M$ is superior to creatinine as a marker of sickle cell nephropathy. However, caution should be applied in patients with haematological malignancies and chronic inflammatory diseases.

Keywords: Sickle cell nephropathy, $\beta_2 M$, Creatine, estimated GFRCr

I. Introduction

Sickle cell disease is a genetic disease due to the inheritance of a sickle cell gene either in the homzygous state or in combination with another haemoglobinopathy such as thalassaemia and Hb C among others.¹ The disease is characterised by acute (crisis) and chronic complications involving multiple organ systems including the renal system. Renal complications of SCD are referred to sickle cell nephropathy (SCN). Renal complications contribute to a significant cause of morbidity and mortality in patients with SCD.²

The burden increases with age and it is estimated that 60% of SCD patients above 45years have microalbuminuria a marker of early renal damage; with 4 - 12% developing end stage renal disease^{2,3}.Platt et al⁴ reported that almost 9% of SCD patients who died due to SCDcomplications manifested overt renal failure.This was identified as a major risk factor for early mortality in adult patients with SCD.

The underlying mechanisms include chronic hypoxia, medullary ischemia, and increased prostaglandin secretion that lead to glomerularhypertrophy, increased glomerular permeability, and proteinuria, which eventually result in chronic renal failure.⁵Furthermore, the hyperosmolar milieu of the medulla, favors hemoglobin S (HbS) polymerization andresults in increased blood viscosity within the medullarcapillaries, leading to loss of concentrating capacity, urinaryacidification, and decreased potassium excretion.⁶ Thus, SCD results in both glomerular and tubular dysfunction which predates chronic renal failure.

The importance of the detection of early features and appropriate management of adverse changes in the renal system of SCD patients cannot be underestimated. Traditional methods of measuring renal impairment using creatinine has been widely critisized and adjudge to have serious limitations.⁷ Its lack of sensivity to detecting early renal changes, subjectivity to spectral interference, dependence on gender and muscle mass necessitates the need for the use of other markers such as β_2 -microglobulin a superior marker for the detection of renal impairment.

 β_2 -Microglobulin (β_2 M), a low molecular weight protein measuring 11,800D. Ithas been identified as the light chain of the class I major histocompatibility antigens. Theyare found on the cell surface of all nucleated cells by which they are synthesized.⁸ β_2 Mis freely filtered in the glomerulus, totally reabsorbed and degraded in the renal tubules. Thus, it is a sensitive marker of the glomerular filtration capacity of the kidney. It is almost absent in urine. Urinary detection is also suggestive of tubular dysfuction.

The burden of SCN in Nigeria is enormous.^{9, 10} However, there are paucity of studies on early markers of renal impairment such as β_2 Min our environment hence this study was undertaken to evaluate the use β_2 Min detecting renal changes in sickle cell patients in Benin City, Nigeria.

This study seeks to compare serum β_2 Mto serum creatinine as markers of renal damage and to correlate β_2 Mwith urea, creatinine, Cr based estimate glomerular filteration rate (eGFRCr)

II. Methodology

This is a cross sectional study conducted at the University of Benin Teaching Hospital Benin City, Nigeria. Eighty three Adult SCD patients in stable disease state (absence of major crises for a period of 3 months) were recruited consecutively from the Adult Haematology Outpatient Unit of the hospital. Their haemoglobin genotypes were determined by Hb electrophore is in alkaline medium. Patients with established end stage renal disease on dialysis, those with haematological malignancies were excluded. Forty healthy volunteers were recruited as controls from the general population.

Participants personal data were obtained using an interviewer administered questionaire. Their height and weight were measured with a stadiometer and weighing scales respectively. The weight and height measurements were used for the estimation of body mass index (BMI) with the formular: BMI = Weight(Kg)/height(m)².

Thereafter 10 mls of venous blood was collected aseptically from the antecubital vein. Three milliliter (3ml) was dispensed into EDTA specimen bottle for packed cell volume estimation and 7 ml into a plain specimen container and allowed to clot. The clotted sample was centrifuged at 1500g for 15 minutes and serum was extracted and stored at -20° C. The EDTA sample was used for estimation of packed cell volume.

The serum sample was used for determination of Creatinine and $\beta_2 M$ levels. Creatinine was determine usingkinetic Jaffe method while β_2 microglobulin level was determine using ELISA method (Quantikine test kit). eGFR was determined using the Cockcroft-Gault formula:¹¹

(140- Age in years) x Weight (kg)/72 x Creatinine(mg/dL) x 0.85(if Female).

The study was approved by the hospital ethical committee and all participants gave informed consent. The data was analyzed with the statistical package for social sciences (SPSS) version 16. Student T test and chi

square were used to compare parametric and non parametric variables between the SCD patients and controls. Pearson's correlation test was used for the correlation test. P value was set at 0.05.

III. Result

The demographic parameters of the SCD patients and controls are shown in table 1. There is no significant difference in the age and sex distribution between both groups. The mean body mass index is significantly lower in SCD patients than in the controls (p = 0.001).

Variables	SCD (n=83)	Controls (n=40)	P value
	$\overline{X} \pm SD$	$\overline{\pmb{X}} \pm \operatorname{SD}$	
Age (yrs)	27.2 ± 7.6	27.4 ± 9.5	0.768
BMI (Kg/m ²)	17.9±4.8	24.1±3.7	0.001
Sex	Freq (%)	Freq (%)	
Males	40 (48.2)	25 (62.5)	0.136
Females	43 (51.8)	15 (37.5)	

Table 1: Age, body mass index and sex distribution of the study population

The mean packed cell volume (PCV), serum urea, creatinine and β_2M in the SCD patients and controls were $25.0\pm3.9\%$ versus $45.6\pm5.8\%$ (p = 0.001), 17.4 ± 7.5 mg/dl versus 24.0 ± 8.0 mg/dl (0.001), 0.7 ± 0.3 mg/dl versus 0.9 ± 0.3 mg/dl (p = 0.003), 3.3 ± 1.0 versus 2.3 ± 0.7 (p = 0.001) respectively. The eGFRCr was higher in SCD patients than in controls (128.9 ± 8.4 ml/min/1.73m² versus 113.6 ± 11.0 ml/min/1.73m²) but the difference was not statistically significant P = 0.316 (Table 2).

0			
Variables	SCD (n=83)	Controls (n=40)	P value
	$\overline{X} \pm SD$	$\overline{\pmb{X}} \pm \operatorname{SD}$	
PCV (%)	25.0 ± 3.9	45.6 ± 5.8	< 0.001
Urea (mg/dl)	17.4 ± 7.5	24.0 ± 8.0	< 0.001
Creatinine (mg/dl)	0.7 ± 0.3	0.9 ± 0.3	0.003
β2 microglobin (mg/L)	3.3 ± 0.98	2.3 ± 0.7	< 0.001
eGFRCr (ml/min/1.73m ²)	128.9 ± 8.4	113.6 ± 11.0	0.316

Table 3 shows the comparison of urea, Cr and $\beta_2 M$ between males and females within each of the study groups. In SCD the difference in means between Cr and $\beta_2 M$ in male and female patients were not significant

statistically (p = 0.353 and 0.114 respectively). However in the controls males have a significantly higher Cr than females (p = 0.001) but there was no significant difference in the mean $\beta_2 M$ level.

37 11						
Variables	SCD(n=83)			Controls (n=40)		
	M (n=40)	F (n=43)	P value	M (n=25)	F (n=15)	P value
	$\overline{X} \pm SD$	$\overline{X} \pm SD$		$\overline{X} \pm SD$	$\overline{X} \pm SD$	
Urea (mg/dl)	17.8±6.5	17.1±8.5	0.715	25.54±9.4	21.6±4.5	0.189
Creatinine (mg/dl)	0.7±0.3	0.7±0.3	0.934	1.1±0.3	0.7±0.1	0.001
β2 microglobin	3.5±1.1	3.1±0.9	0.114	2.3±0.7	2.3±0.6	0.815
(mg/L)						
eGFRCr	136.4±15.0	122.5±8.0	0.854	109.3±17.0	120.3±9.2	0.637
$(ml/min/1.73m^2)$						

Table 3: Biochemical parameters based on sex distribution of the study population

Using eGFRCr, 21 (25.3%) of the SCD patients and 14 (35%) of the controls had subnormal renal function. The difference in not statistically significant (X2 = 1.247; p = 0.264). However using β_2 M, 64 (77.1%) of the SCD patients had impaired renal function compared to 10 (25.0%) of the controls (X² = 30.58; p = 0.001) as in table 4.

Table 4: Comparison of the proportion of SCD patient and Controls with impaired renal function using eGFR and B_2M

Variables	SCD	Controls	X^2	P value
eGFR Cr				
<90 ml/min/1.73m ²	21 (25.3)	14 (35.0)	1.247	0.264
\geq 90 ml/min/1.73m ²	62 (74.7)	26 (65.0)		
$\beta_2 M$				
≤2.7mg/dl	19 (22.9)	10 (25.0)	30.580	0.001
>2.7mg/dl	64 (77.1)	30 (75.0)		

Table 5 compares the proportion of subjects in each that has renal dysfunction using eGFR and that using $\beta_2 M$. In the SCD group, using $\beta_2 M$ the proportion of renal dysfunction detectable is significantly higher than with eGFR (X² = 48.58; p = 0.000). In the control (non SCD)group, no significant difference was found with use of B₂M and eGFRCr (X² = 0.952; p = 0.329).

	· · · · · · · · · · · · · · · · · · ·			
Variables		SCD		P value
	eGFR Cr	$\Box_2 \mathbf{M}$		
Normal	62 (74.7)	19 (22.9)	44.580	0.000
Abnormal	21 (25.3)	64 (77.1)		
		Controls		
	eGFR Cr	$\square_2 \mathbf{M}$		
Normal	26 (65.0)	30 (75.0)	0.952	0.329
Abnormal	14 (35.0)	10 (25.0)		

 Table 5: Comparison of B2M and eGFRCr as markers of Renal Dysfunction

In the SCD patients, there were statistically significant positive correlations between $\beta_2 M$ and Urea(r = 0.548; p = 0.000) and between $\beta_2 M$ and Cr (0.533; p = 0.000). A negative correlation was established between $\beta_2 M$ and eGFR Cr but it was not statistically significant (r = -0.134; p = 0.222) (Table 6).

Table 6: Correlation between β_2 M, Urea, Cr and eGFR Cr					
		Urea	Creatinine	eGFR Cr	
$\beta_2 M$	R	0.548	0.533	-0.134	
	P value	0.000	0.000	0.222	

Table 6: Correlation between β_2 M, Urea, Cr and eGFR Cr

IV. Discussion

Sickle cell nephropathy (SCN) is a cause of significant morbidity and mortality in patients with sickle cell disease. The need for the use of a sensitive marker for detection of early changes in renal function cannot be overemphasized. Creatinine based assay has several limitations necessitating the need for the use of superior markers for the detection of renal injuries. $\beta_2 M$ is one of such markers capable of replacing Cr assay.

In this study, the mean Cr levels was significantly lower in SCD patients than in controls while eGFR Cr was higher in SCD patients than in controls though not statistically significant. The relatively lower Cr and higher eGFR Cr can be attributed to the lower body mass of the SCD subjects and the associated glomerular hyperfilterationchanges. Hyperfiltration is one of the early parthological changes in SCN. However, the B_2M was significantly higher in the SCD group.

Sessoet al^{12} in a comparative study on renal function in homozygous SS and heterozygous AS subjects reported a significantly increased $\beta_2 M$ in patients with SS. Similarly they observed a significantly lower serum creatinine and a significantly higher eGFR. In another related study, de Jong et al,¹³ reported a significantly elevated $\beta_2 M$ in SCD compared to controls. Although measurement of Cr issimple and easily available, it is reported that almost 50% of patients withimpaired GFR have normal Cr¹⁴. This suggests that $\beta_2 M$ is a better marker of renal derangement thanthe eGFRCr in SCD.

The proportion of SCD patients with subnormal eGFRCr is 25.3% compared to 77.1% using $\beta_2 M$ estimation. This difference is statistically significant. Thus $\beta_2 M$ is superior to eGFRCr in detecting renal dysfunction. This is similar to the observations of Voskaridouet al¹⁵ however, values observed in the index study were relatively higher than theirs.

Cr based eGFR is established to be dependent on age, body mass, sex and it is prone to interferences unlike $\beta_2 M$. This study has shown that sex differences did not affect $\beta_2 M$ levels both in the SCD and control groups. Guido et al¹⁶ demonstrated the age independence of $\beta_2 M$ making it a suitable marker for use in evaluating glomerular dysfunction across all age groups. $\beta_2 M$ has been observed to become elevated by the age of 20 years while significant observable deterioration in GFR becomes obvious by the age of 35 yrs in sickle cell patients.¹⁷ Using $\beta_2 M$ as marker of renal dysfunctionin SCD patients, SCN will be detected earlier and appropriate intervention initiated compared to the use of the traditional eGFRCr estimation.

In conclusion, it is obvious that the use of $\beta_2 M$ to evaluate renal function in SCD is superior to creatinine and eGFRCr. However, caution should be exercised in interpreting results of patients with haematological malignancies and chronic inflammatory conditions as connective tissue disease as its level will be elevated in these conditions.

References

- [1]. Asnani MR. 2010. Sickle Cell Disease. In: JH Stone, M Blouin, editors. International Encyclopedia of Rehabilitation. Available online: http://cirrie.buffalo.edu/encyclopedia/en/article/252/
- [2]. Guasch A, Navarrete J, Nass K, Zayas CF. Glomerular involvement in adults with sickle cell hemoglobinopathies: prevalence and clinical correlates of progressive renal failure. Journal of the American Society of Nephrology 2006; 17, 2228–2235.
- [3]. Powars DR, Elliott-Mills DD, ChanL, Niland J, Hiti AL, Opas LM Johnson C. Chronic renal failure in sickle cell disease: risk factors, clinical course, and mortality. Annals of Internal Medicine 1991; 115, 614–620.
- [4]. Platt OS, Brambilla DJ, Rosse WF et al. Mortality in sickle cell disease. Life expectancy and risk factors for early death. N Engl J Med 1994; 330: 1639–1644.
- [5]. Wesson DE. The initiation and progression of sickle cell nephropathy. Kidney International 2002; 61: 2277–2286
- [6]. Sharpe CC, Thein SL. Sicke cell nephropathy a practical approach. Br J Haematol. 2011;155(3): 287-297.
- [7]. National Kidney Foundation-K/DOQI. Clinical Practice Guidelines for Chronic Kidney Disease, Evaluation, Classification and Stratification.Am J kidney Dis.2002; 39 (Suppl 1):S1-S266.
- [8]. Creswell P, Springer T, Strominger JL, Turner MJ, Grey HM, Kubo RT: Immunological identity of the small subunit of HLA antigens and β₂-microglobulin and its turnover on the cell membrane. Proc Nat Acad Sci 1974; 71:2123 2207.
- [9]. Aneke JC, Agegoke AO, Oyekunle AA, Osho PO, Sanusi AA, Okocha EO et al. Degrees of kidney disease in Nigerian Adults with sickle cell disease. Med Princ Pract 2014; 23: 271 274.
 [10]. Bolarinwa RA, Akinlade KS, Kuti M, et al: Renal disease in adult Nigerians with sickle cell anemia: a report of prevalence, clinical
- [10]. Bolarinwa KA, Akiniade KS, Kuti M, et al: Renal disease in adult Nigerians with sickle cell anemia: a report of prevalence, clinical features and risk factors. Saudi J Kidney Dis Transpl 2012; 23: 171–175.
- [11]. Cockroft D, Gault MK. Prediction of creatinine clearance from serum creatinine. Nephrol 1976; 16: 31-41.
- [12]. Sesso R, Almeida MA, Figueiredo MS, Bordin JO. Renal dysfunction in patients with sickle cell anemia or sickle cell trait. Brazillian Journal of Medical and Biological Research 1998: 31: 1257-1262.
- [13]. de Jong PE, de Jong-van den Berg LT, Sewrajsingh GS, Schouten H, Donker AJ, Statius van Eps LW. Beta-2-microglobulin in sickle cell anaemia. Evidence of increased tubular reabsorption. Nephron. 1981;29(3-4):138-41.
- [14]. Perrone R, Madias N, Levey A. Serum creatinine as an index of renal function: new insights into old concepts. Cin Chem 1992; 38:1933–1953.
- [15]. Voskaridou E, Terpos E, Michail S, Hantzi E, Anagnostopoulos A, Margeli A et al. Early markers of renal dysfunction in patients with sickle cell/β-thalassemia. Kidney International 2006; 69: 2037–2042
- [16]. Guido F, IrisW,1 Friedrich P,Jochen HHE and Klaus J. Are Cystatin C and B2-Microglobulin better markers than serum Creatinine for prediction of a normal glomerular filtration rate in pediatric subjects? Clinical Chemistry 1997; 43(6): 1077-1078.
- [17]. Katis R in Scheinman JI; Tools to detect and modify sickle cell nephropathy. Kidney International 2006: 69: 1927–1930.